



Univerzita Komenského v Bratislave
Fakulta matematiky, fyziky a informatiky



Michal Bodík

Autoreferát dizertačnej práce

Exfoliácia a pokročilá funkcionálna dvoj-dimenzionálnych materiálov

na získanie akademického titulu philosophiae doctor

v odbore doktorandského štúdia:

4.1.4. Kvantová elektronika a optika

**Miesto a dátum:
Bratislava, 2020**

Dizertačná práca bola vypracovaná v dennej forme doktorandského štúdia

na Fyzikálnom ústave Slovenskej akadémie vied

Predkladateľ: **Michal Bodík**
Fyzikálny ústav, SAV
Dúbravská cesta 9
845 11 Bratislava

Školiteľ: **Dr. Rer. Nat Peter Šiffalovič**
Fyzikálny ústav SAV

4.1.4. Kvantová elektronika a optika

Predseda odborovej komisie:

.....
(meno a priezvisko s uvedením titulov a hodností
a presná adresa jeho zamestnávateľa)

Abstract

The cancer treatment is a complicated process build-up from a large number of specific procedures and methods. Because of this, it is necessary to focus on the multidisciplinary research of new alternative approaches, technologies, and materials. Targeted cancer treatment using low-dimensional materials joints multiple scientific disciplines on the edge of physics, chemistry, and biology with the target on the preparation of new drug nanocarriers. A novel family of two-dimensional materials allows unprecedented possibilities in the area of bioconjugation thanks to the high surface-to-volume ratio. In this thesis, I focused on the liquid-phase exfoliation of two candidates for the drug nanocarriers, namely MoS₂ and graphene oxide. I studied the exfoliation process of those two-dimensional materials, resulting size of nanoplatfoms and their potential for bioconjugation. The second step was the study of the possible functionalization routes of developed nanoplatfoms. For each of the two, I developed a specific procedure for attaching the specific antibody to its surface.

Introduction

Research of novel materials always had a firm place among other fields of physics. Their discovery opens new possibilities in all the fields of everyday life, whether its new technologies, biomedical applications, or simply basic research.

One such breakthrough was when graphene was first observed in 2004 [1]. It was the first-ever two-dimensional (2D) material observed and opened a new chapter in the material research [2]–[5]. Graphene is a single layer of carbon atoms, crystallized in a hexagonal structure. It has unique mechanical [6], electrical [7], [8], thermal [9], [10] and optical [11] properties. The discovery of graphene led to the discovery of a whole new family of 2D materials [11]–[13]. Each material has unique properties and is suitable for different applications in different fields. The largest family of 2D materials are transition metal dichalcogenides [12], [14]. Nanomaterials from this family contain one atom of transition metal (M) sandwiched between two atoms of chalcogenide (X), thus forming an MX_2 structure. Such a crystalline structure allows the exfoliation to monolayers, similar to the graphene.

One such promising application is in the drug delivery systems [15]. The big advantage of 2D materials is their large specific surface. In the case of graphene, all its atoms are at its surface. The 2D nanomaterial can serve as the smallest possible nanocarrier, that carries the drug into the cells.

However, the functionalization of 2D nanomaterials has not a universal procedure. The same drug will require a different binding procedure if bound with graphene or transition metal dichalcogenide.

In my thesis, I focused on the production, physicochemical characterization, and functionalization of 2D materials for bioapplications. Namely, I focused on graphene oxide and MoS_2 and their use in targeted cancer treatment. I developed a reliable exfoliation method for both of the nanomaterials, which will result in the exfoliated 2D nanomaterial in the solution. The complete physicochemical characterization is presented as a solid background for bioconjugation with a specific antibody. The antibody M75 was used for its specificity towards CAIX antigen, which is expressed on the cancer cell's surface.

1. Two-dimensional materials in cancer treatment

1.1. Graphene Oxide

Graphene oxide (GO) is a successor of the first two-dimensional (2D) material, i.e. graphene. GO is obtained from graphite by oxidation and exfoliation procedure [1], [2]. Oxygen groups increase the interlayer distance and successively decrease the strength of van der Waals bonding between the neighboring graphite sheets. Followingly, the oxidized graphite is easier to exfoliate to monolayers and what is crucial to our application, it is soluble in water. The schematics of the oxidation and exfoliation process is shown in Fig. 1.

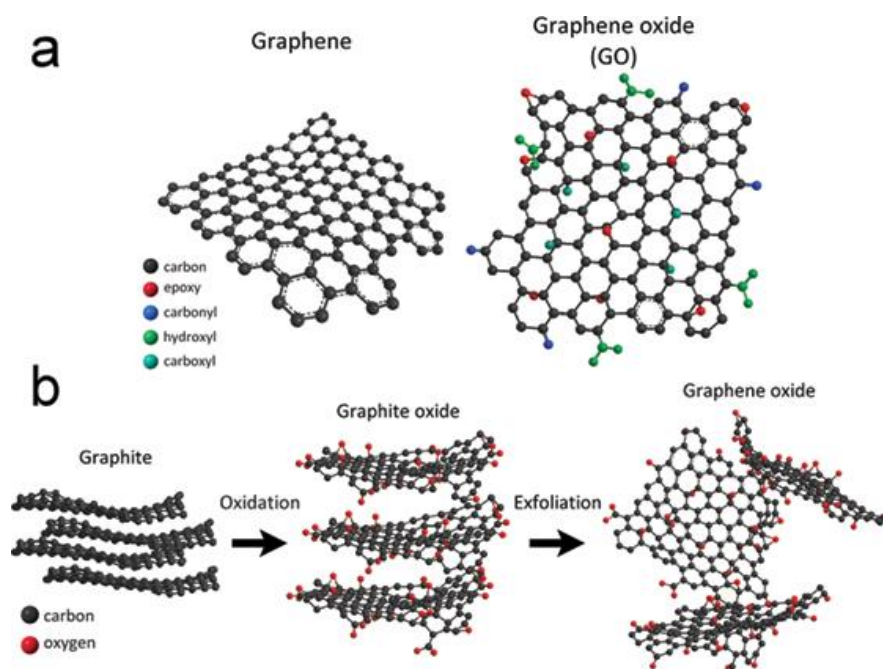


Fig. 1: a) Schematic representation of the GO structure and b) the production of the monolayer GO process[3].

Oxygen groups are used for bioconjugation. Mainly, the carboxyl groups are used for bioconjugation because of the strong covalent bonding between carboxyl and amine groups [4]–[6]. The linking between carboxyl and amine group is facilitated by crosslinking agents EDC (1-ethyl-3-(3-dimethylaminopropyl)carbodiimide hydrochloride) and sulfo-NHS (N-hydroxysulfosuccinimide) [5], [6]. These two crosslinkers mediate the formation of a covalent bond and are present only during the formation of the amide bond between carboxyl and amine. After the amide bond is created, the crosslinkers EDC and sulfo-NHS are no longer active, as they are quickly hydrolyzed in the water.

1.2. Molybdenum disulfide

Molybdenum disulfide is a transition metal dichalcogenide, that can be exfoliated to monolayers [7]. Contrasting to graphene, bulk MoS₂ is an indirect semiconductor with a bandgap of 1.2 eV, while the monolayer has a direct bandgap of 1.9 eV due to the quantum confinement effect [8], [9]. The photoluminescence (PL) of monolayer MoS₂ is located at approximately 650 nm so that the red PL can be observed in MoS₂ monolayer [8], [9]. The presence of PL and the metal atoms in the chemical structure permits the MoS₂ detectability in soft tissues.

The functionalization of MoS₂ is more complicated than in the case of GO. The lack of functional groups prevents the use of conventional bioconjugation techniques. Instead, the sulfur vacancies are utilized to functionalize the MoS₂ [10]–[12]. The defect site will attach a sulfur atom and thus promote the bonding. This sulfur atom can be present either in a thiol group [13]–[15] or sulfur from lipoic acid [11], [16], [17]. The linkage between MoS₂ nanosheet and the conjugate is often provided by the PEG [10], [15], [16], polyethyleneimine [17] or different thiolated ligand [13]. PEG is a well-known excipient [18], that is often used in medicine to increase the stability in the physiological condition and decrease the reaggregation and immunogenicity [19], [20] of the drugs. The other end of the PEG molecule is not attached to MoS₂ nanosheet and can be used for the attachment of a drug [16] (Fig. 3), toxin [15], or an aptamer for tracking of processes inside the cells [17].

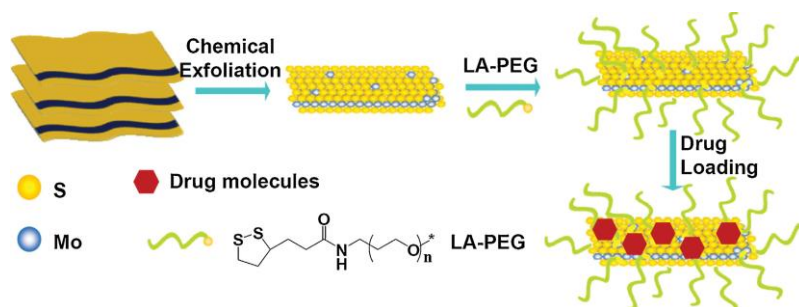


Fig. 3: A schematic representation of MoS₂ functionalization via lipoic acid [16].

2. Langmuir films

Originally, a Langmuir film is a self-assembled layer of molecules on the water surface [21]. The Langmuir films formation consists of the dropping of a solution of lipid molecules on the surface of the water subphase. The amphiphilic molecules spread across the subphase surface, and the solvent evaporates. It leaves the molecules mobile on the surface, and a pair of movable barriers slowly confine the area, where the molecules can float. The barriers movement forces the molecules on the surface to compress and form a monolayer film of the oriented molecules with their polar ends in the water subphase.

2.1. Langmuir films of 2D materials

2.1.1. Graphene oxide

It is not straightforward to produce Langmuir films from GO on the water subphase, because of its amphiphilicity. There are several ways to overcome the dispersibility of GO in the water subphase.

We can use the amphiphilicity to work against the dispersibility in the water subphase [22]. By increasing the pH of the water, the carboxyl groups at the edge of GO flakes are deprotonated. Therefore, the GO flakes get more charged, and the solubility in water increases. On the other hand, if we decrease the pH of the subphase, the GO flakes will become insoluble in the water subphase. Plus, good spreadability of our solution on the water surface is guaranteed by adding methanol to the GO dispersion. Methanol will not change the stability of the dispersion of GO, but its large difference in the surface tension will cause the droplet to spread across the water and not to sink in.

2.1.2. MoS₂

Making Langmuir films of MoS₂ requires similar condition as GO. The MoS₂ can be directly exfoliated in ethanol/water mixture, so the good spreading is guaranteed. And because the MoS₂ is to some extent dispersible in water, changing the pH to lower values is advisable as well.

3. Aim of the work

The aim of this thesis is to study the preparation, characterization, and applications of two-dimensional materials.

Namely, the liquid-phase exfoliation of MoS₂ and GO to produce sufficiently small 2D nanoplateforms, that will be used as nanocarriers for specific antibodies.

In my thesis, I focused on the liquid-phase exfoliation of MoS₂ and GO using ultrasound. The basic optimization of the parameters of exfoliation was studied to maximize the yield of the desired size of nanoplateforms. Further size-selective sorting was performed using a centrifuge.

The dispersions of nanoplateforms were transferred onto substrates using the Langmuir-Schaefer approach before analysis.

The resulting size distribution of nanoplateforms was detected by imaging methods such as AFM, SEM or TEM and supported by spectroscopy methods such as Raman spectroscopy or absorption spectroscopy.

Nanoplateforms with desired size distributions were chemically functionalized to enable the bioconjugation of antibodies.

4. Analytical techniques for Langmuir films

The monitoring of the surface pressure is essential for the Langmuir film formation. The change in the surface pressure of the subphase is the primary information on the forming layer during the compression. In the case of low-dimensional nanomaterials, the main interest is in the formation of a closed, homogenous monolayer suitable for further applications. Here, we define the monolayer as a film, which has the highest possible surface density of given nanomaterial. The highest density is achieved when the nanomaterial is compressed to a critical pressure π_c . If the film is compressed above the π_c , the monolayer collapses what promotes the nucleation of the second layer. The film's elastic modulus $E = -A \frac{d\pi}{dA}$ or its inverse, the film's compressibility C , can be employed to assess the critical pressure π_c . A numerical calculation of the Elastic modulus E from the π - A isotherm is utilized to determine the π_c correctly [23], [24].

In general, imaging null ellipsometry can be used to track the formation of the molecular Langmuir films with micrometer spatial resolution [25]. Its wide-spread modification, the Brewster angle microscopy (BAM), uses the p -polarized light incident at the Brewster angle, which is close to 53° for the water subphase. The large difference between the optical refraction index of many low-dimensional nanomaterials and water predetermines the BAM as the method of the first choice for studying the compression of the nanomaterial films on the water subphase [26]–[33].

The X-ray scattering techniques belong to the advanced *in-situ* characterization methods applicable to the formation of Langmuir films. In a typical GISAXS experiment, an X-ray beam impinges onto the water surface at a grazing-incidence angle close to the critical angle for the total reflection. Each nanoobject in the Langmuir film acts as a scattering center and produces a partially scattered wave. All partially scattered waves interfere and are recorded in the far-field as the intensity GISAXS scattering pattern [23], [34]. By evaluating the GISAXS pattern, we can monitor the interparticle distance and surface coverage.

5. Analytical techniques

The resolution of optical microscopy is not sufficient for the surface characterization of nanomaterials. The resolution limit is given by the Rayleigh criterion $d = \lambda/2NA$, where λ is the light wavelength, and NA is the numerical aperture of the used objective. For visible light, the typical resolution limit is approximately 200 nm. Consequently, there is no possibility to distinguish between the two neighboring nanoparticles if their dimensions are in the range of tens of nanometers.

The AFM technique measures the morphology of the sample surface by evaluating the forces acting between the vibrating probe and surface. The dimensions of the probing tip limit the AFM resolution. Within today's technology, the sharpness of the scanning tip can be less than a few nanometers.

The absorption spectroscopy measures the optical response in the UV-Vis-NIR range. The change in the electronic structure of material results in the change of the absorption spectra. The electrons and their energy levels are responsible for the energy of absorbed or emitted light. On top of this, the concentration of the nanomaterial in the solution can be calculated from Beer-Lambert law.

The Raman spectroscopy was employed to reveal the crystallinity of prepared nanomaterials. In Raman spectroscopy, the sample is irradiated with a high-intensity laser, and the inelastically scattered light is detected. The photons after inelastic scattering carry the information about the vibrations of the atoms in crystals or molecules in the sample.

The X-ray photoelectron spectroscopy (XPS) was used to analyze the atomic composition of samples. The XPS analyzes the kinetic energy of photoelectrons emitted from the sample surface after the absorption of X-ray photons. The binding energy can be determined as the difference between the X-ray photon energy and the kinetic energy of emitted photoelectrons.

The resolution limit of the optical microscope can be surpassed by decreasing the wavelength of the light. The electron microscopes use this fact, and instead of the visible light, they use electrons as the probing particles. The microscope resolution is given by the de Broglie wavelength of used electrons, and hence the atomic resolution can be achieved. The transmission electron microscope (TEM) uses electromagnetic lenses to create a magnified image of a thin sample on a two-dimensional detector.

6. Results

6.1. MoS_2

The successful exfoliation of MoS_2 is crucial for the functionalization. On top of that, the highest possible concentration of exfoliated MoS_2 is desired. However, during the exfoliation of MoS_2 in ethanol/water mixtures, the elevated concentration will cause the oxidation of MoS_2 instead of the exfoliation (Fig. 4).

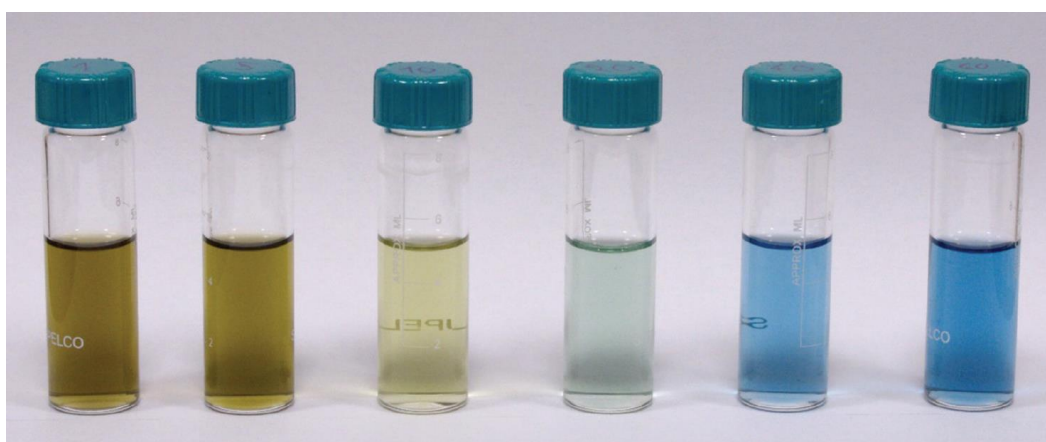


Fig. 4: The photographs of the supernatants after MoS_2 exfoliation and centrifugation for the initial concentrations: 1, 5, 10, 20, 40 and 60 mg/ml (from left to right).

Here, the water and dissolved oxygen act as the oxidizing agent during liquid-phase exfoliation. Along with that, the oxidation is accompanied by the release of SO_4^{2-} and H^+ that will cause the pH of the dispersion to drop. The higher the initial concentration of MoS_2 in the dispersion, the more the pH drops. After the pH dropped over the stability point of the MoS_2 colloidal solution, all the exfoliated crystals start to sediment. Furthermore, only the MoO_x nanoparticles remain the form of a colloidal solution.

Knowing the limitations of the liquid-phase exfoliation of MoS_2 , the highest concentration of MoS_2 obtainable by this method was discovered.

Langmuir films of MoS_2 were prepared for the characterization of the MoS_2 nanosheets. The average thickness of MoS_2 was determined to be four layers and the lateral size of nanosheets was found to be in the range of 30-70 nm.

The bioconjugation of MoS_2 was performed via so-called biotin-avidin-biotin bridge (Fig. 5). First, the MoS_2 was functionalized by the lipoic acid-PEG-biotin molecules.

Secondly, avidin reacts with the terminal biotin and leaves three empty reaction sites for extra biotin molecules. These empty reaction sites react with the biotinylated antibodies to finish the process.

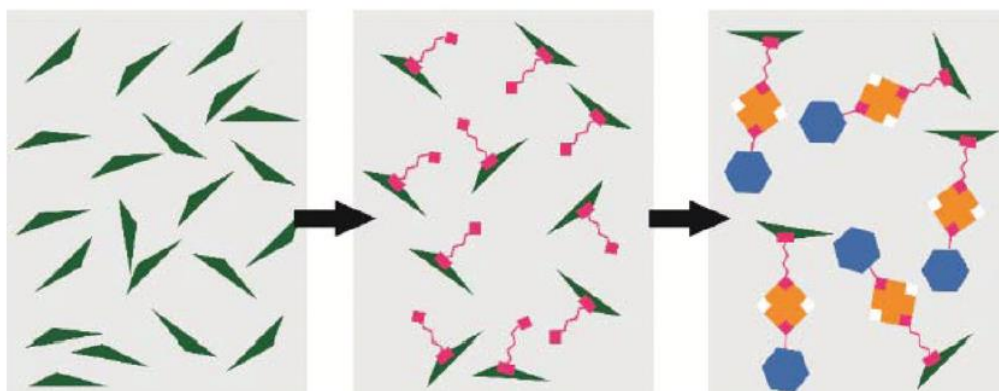


Fig. 5: Schematics of the three-step functionalization workflow: first, the MoS₂ nanosheets are exfoliated with LPE in deionized water. In the second step, PEGylation proceeds and in the last step, the M75 antibody is conjugated via a biotin–avidin–biotin bridge to the nanoplatform.

6.2. Graphene oxide

The main advantage of GO, when compared to MoS₂, is the ability to control its size. Monolayer GO with different lateral sizes ranging from micrometers to tens of nanometers can be prepared by carefully selecting the centrifugation steps.

In our work, the antibody was bonded to magnetic nanoparticles. Magnetic nanoparticles could be further used for MRI imaging or the killing of the cancer cell by hyperthermia. The magnetic nanoparticles were terminated by amino group and the bonding process relies on the formation of an amide bond between the nanoparticle and GO sheet.

First, a PBS buffer solution containing a small amount of TWEEN®20 was mixed with GO solution. TWEEN is a surfactant that opposes the agglomeration of GO during the functionalization. Second, EDC and sulfo-NHS were added to the solution. EDC and sulfo-EDC are zero-length cross-linkers, that promote the formation of an amide bond. Furthermore, in the last step, the antibody-containing magnetic nanoparticles were added. This way, the amide bond between the carboxyl group on GO sheet and amine group on the magnetic nanoparticle was created (Fig. 6).

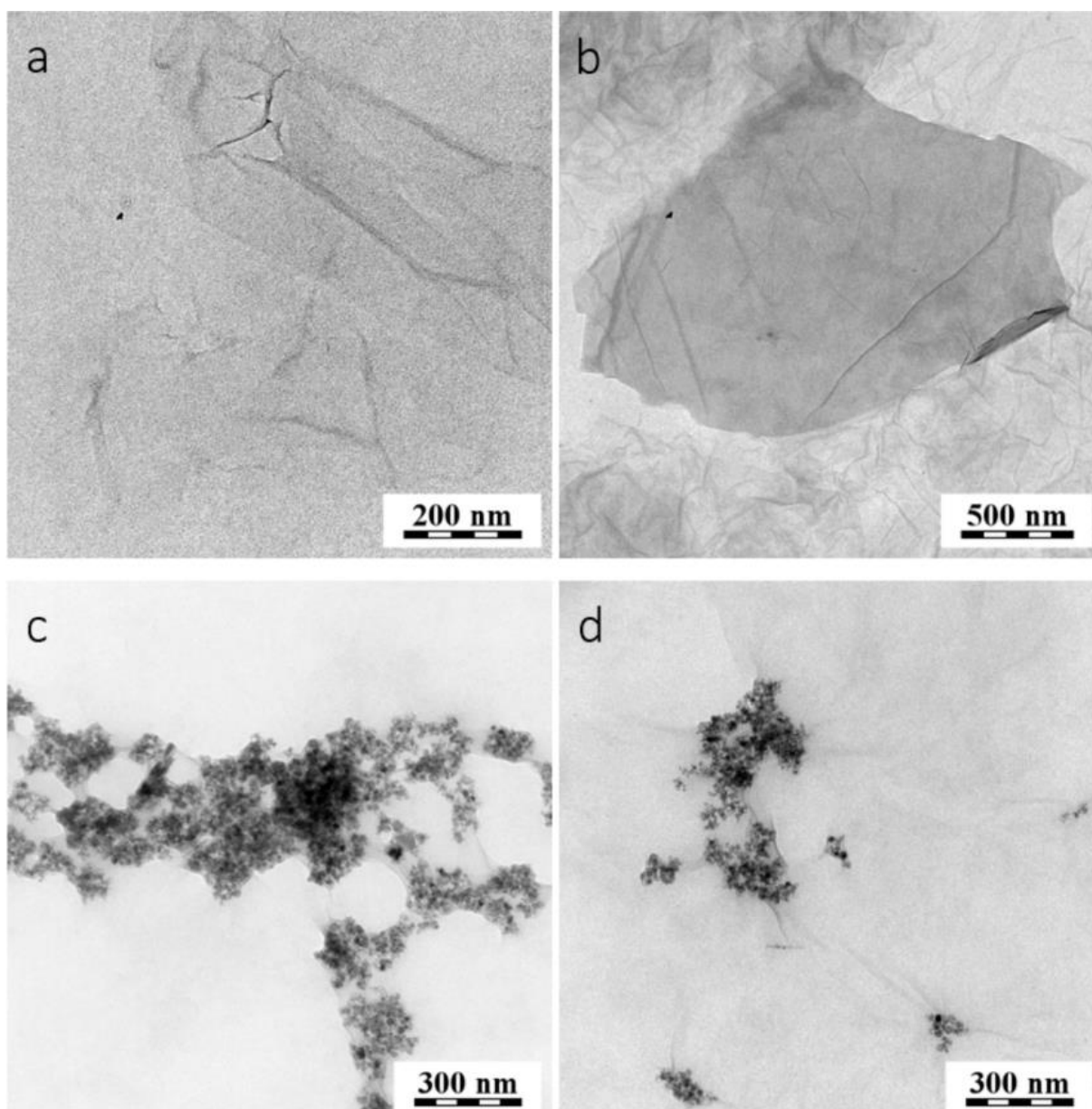


Fig. 6: Transmission electron microscopy (TEM) micrographs of the (a,b) pure GO platforms; (c) graphene oxide platforms with magnetic nanoparticles modified antibodies, and (d) graphene oxide platforms with magnetic nanoparticles modified antibodies in the presence of cross-linkers.

Conclusions

The aim of this thesis was to prepare a functionalized 2D nanoplateforms, that can be used for targeted cancer treatment. In this work, we focused on the production of 2D nanoplateforms using the liquid-phase exfoliation method.

The powders of MoS₂ or GO were dispersed in an appropriate solvent and exfoliated using ultrasonic waves. The resulting dispersions of exfoliated 2D materials were centrifuged to remove the unexfoliated material. The supernatant contains exfoliated MoS₂ nanoplateforms and the precipitate unexfoliated MoS₂ crystals.

After the dispersions of 2D materials were prepared, the size-distribution and their crystallinity were characterized. Using Langmuir-Schaefer deposition, both MoS₂ and GO were transferred onto silicon or glass substrate. The characterization of MoS₂ and GO nanoplateforms was performed using AFM, Raman spectroscopy, SEM, TEM, XPS, and absorption spectroscopy. In this step, we analyzed whether the nanoplateforms are of the desired size and are suitable for functionalization.

The functionalization of MoS₂ relies on the presence of the sulfur vacancies on its surface. The lipioic-acid modified PEG binds to the MoS₂ surface and leaves the second end of the PEG chain ready for further functionalization. In our work, we used biotin as a terminating agent. The terminating biotin binds in the next step with avidin, which has four binding sites for biotin. In the last step, the biotinylated antibody binds with avidin and closes the biotin-avidin-biotin bridge.

On the other hand, GO has already on its surface functional oxygen groups. In our work, we used carboxyl groups on the GO to directly bind GO with antibody-coated magnetic nanoparticles. We used coupling agents such as EDC and sulfo-NHS to promote the formation of the amide bond between GO and the magnetic nanoparticle.

The functionality of such nanoplateforms was further studied on the cancer cells.

The final products of this work are two types of nanoplateforms for targeted cancer treatment. One of the nanoplateforms is based on GO, i.e., carbon-based nanomaterial and the second one is based on inorganic nanomaterial MoS₂.

Publication presented in the dissertation thesis

- ELIÁŠOVÁ SOHOVÁ, Marianna - **BODIK, Michal** - ŠIFFALOVIČ, Peter** - BUGÁROVÁ, Nikola - LABUDOVÁ, Martina - ZAŤOVIČOVÁ, Miriam - HIANIK, Tibor - OMASTOVÁ, Mária - MAJKOVÁ, Eva - JERGEL, Matej - PASTOREKOVÁ, Silvia. Label-free tracking of nanosized graphene oxide cellular uptake by confocal Raman microscopy. In *Analyst*, 2018, vol. 143, iss. 15, p. 3686-3692. ISSN 0003-2654.
 - Cited in: Kim, Mina, et al. „Graphene oxide-induced neurotoxicity on neurotransmitters, AFD neurons and locomotive behavior in *Caenorhabditis elegans*.“ *NeuroToxicology* 77 (2020): 30-39.

Piperno, Anna, et al. “Casting Light on Intracellular Tracking of a New Functional Graphene-Based MicroRNA Delivery System by FLIM and Raman Imaging.” *ACS applied materials & interfaces* 11.49 (2019): 46101-46111.

Kim, Jongwoo, et al. "SERS-based particle tracking and molecular imaging in live cells: toward the monitoring of intracellular dynamics." *Nanoscale* 11.45 (2019): 21724-21727.

Zaharie-Butucel, Diana, et al. "Efficient combined near-infrared-triggered therapy: phototherapy over chemotherapy in chitosan-reduced graphene oxide-IR820 dye-doxorubicin nanoplateforms." *Journal of colloid and interface science* 552 (2019): 218-229.
- KÁLOSI, Anna - DEMYDENKO, Maksym - **BODIK, Michal** - HAGARA, Jakub - KOTLÁR, Mário - KOSTIUK, Dmytro - HALAHOVETS, Yuriy - VÉGSO, Karol - ROLDAN, Alicia Marin - MAURYA, Gulab Singh - ANGUS, Michal - VEIS, Pavel - JERGEL, Matej - MAJKOVÁ, Eva - ŠIFFALOVIČ, Peter**. Tailored Langmuir–Schaefer Deposition of Few-Layer MoS₂ Nanosheet Films for Electronic Applications. In *Langmuir*, 2019, vol. 35, no. 30, p. 9802-9808. ISSN 0743-7463.
 - Cited in: Kazak, Alexandr V., et al. "Floating layers and thin films of mesogenic mix-substituted phthalocyanine holmium complex." *Thin Solid Films* (2020): 137952.
- BUGÁROVÁ, Nikola** - ŠPITÁLSKY, Zdenko - MIČUŠÍK, Matej - **BODIK, Michal** - ŠIFFALOVIČ, Peter - KONERACKÁ, Martina - ZÁVIŠOVÁ, Vlasta - KUBOVČÍKOVÁ, Martina - KAJANOVÁ, Ivana - ZAŤOVIČOVÁ, Miriam - PASTOREKOVÁ, Silvia - ŠLOUF, Miroslav - MAJKOVÁ, Eva - OMASTOVÁ, Mária**. A multifunctional graphene oxide platform for targeting cancer. In *Cancers*, 2019, vol. 11, no. 6, art. no. 753, [19] p. ISSN 2072-6694.

- **BODIK, Michal**** - ANNUŠOVÁ, Adriana - HAGARA, Jakub - MIČUŠÍK, Matej - OMASTOVÁ, Mária - KOTLÁR, Mário - CHLPÍK, Juraj - CIRÁK, Július - ŠVAJDLENKOVÁ, Helena - ANGUŠ, Michal - ROLDÁN, Alicia Marín - VEIS, Pavel - JERGEL, Matej - MAJKOVÁ, Eva - ŠIFFALOVIČ, Peter. An elevated concentration of MoS₂ lowers the efficacy of liquid-phase exfoliation and triggers the production of MoO_x nanoparticles. In *Physical Chemistry Chemical Physics*, 2019, vol. 21, no. 23, p. 12396-12405.
- KALOSI, Anna – LABUDOVA, Martina – ANNUSOVA, Adriana – BENKOVICOVA, Monika – **BODIK, Michal** – KOLLAR, Jozef – Kotlar, Mario – KASAK, Peter – JERGEL, Matej – PASTOREKOVA, Silvia – SIFFALOVIC, Peter – MAJKOVA, Eva. A bioconjugated MoS₂ based nanoplatform with increased binding efficiency to cancer cells. In *Biomaterials Science*, 2020, vol. 7, no. 8, p. 1973-1980

References

- [1] W. S. Hummers and R. E. Offeman, "Preparation of graphitic oxide," *J. Am. Chem. Soc.*, vol. 80, no. 6, p. 1339, 1958.
- [2] D. C. Marcano *et al.*, "Improved synthesis of graphene oxide," *ACS Nano*, vol. 4, no. 8, pp. 4806–4814, Aug. 2010.
- [3] E. J. Amieva, J. López-Barroso, A. L. Martínez-Hernández, and C. Velasco-Santos, "Graphene-Based Materials Functionalization with Natural Polymeric Biomolecules," *Recent Adv. Graphene Res.*, pp. 257–298, 2016.
- [4] R. Imani, S. H. Emami, and S. Faghihi, "Nano-graphene oxide carboxylation for efficient bioconjugation applications: a quantitative optimization approach," *J. Nanoparticle Res.*, vol. 17, no. 2, pp. 1–15, Feb. 2015.
- [5] C. Shan, H. Yang, D. Han, Q. Zhang, A. Ivaska, and L. Niu, "Water-soluble graphene covalently functionalized by biocompatible poly-L-lysine," *Langmuir*, vol. 25, no. 20, pp. 12030–12033, Oct. 2009.
- [6] G. T. Hermanson, *Bioconjugate techniques*. Academic press, 2013.
- [7] K. F. Mak, C. Lee, J. Hone, J. Shan, and T. F. Heinz, "Atomically thin MoS₂: A new direct-gap semiconductor," *Phys. Rev. Lett.*, vol. 105, no. 13, p. 136805, Sep. 2010.
- [8] A. Splendiani *et al.*, "Emerging photoluminescence in monolayer MoS₂," *Nano Lett.*, vol. 10, no. 4, pp. 1271–1275, Apr. 2010.
- [9] G. Eda, H. Yamaguchi, D. Voiry, T. Fujita, M. Chen, and M. Chhowalla, "Photoluminescence from chemically exfoliated MoS₂," *Nano Lett.*, vol. 11, no. 12, pp. 5111–5116, Dec. 2011.
- [10] S. S. Chou *et al.*, "Ligand conjugation of chemically exfoliated MoS₂," *J. Am. Chem. Soc.*, vol. 135, no. 12, pp. 4584–4587, Mar. 2013.
- [11] T. Liu *et al.*, "Iron oxide decorated MoS₂ nanosheets with double PEGylation for chelator-free radiolabeling and multimodal imaging guided photothermal therapy," *ACS Nano*, vol. 9, no. 1, pp. 950–960, Jan. 2015.
- [12] M. Makarova, Y. Okawa, and M. Aono, "Selective adsorption of thiol molecules at sulfur vacancies on MoS₂(0001), followed by vacancy repair via S-C dissociation," *J. Phys. Chem. C*, vol. 116, no. 42, pp. 22411–22416, Oct. 2012.

- [13] J. S. Kim, H. W. Yoo, H. O. Choi, and H. T. Jung, “Tunable volatile organic compounds sensor by using thiolated ligand conjugation on MoS₂,” *Nano Lett.*, vol. 14, no. 10, pp. 5941–5947, Oct. 2014.
- [14] X. Chen, N. C. Berner, C. Backes, G. S. Duesberg, and A. R. McDonald, “Functionalization of Two-Dimensional MoS₂: On the Reaction between MoS₂ and Organic Thiols,” *Angew. Chemie - Int. Ed.*, vol. 55, no. 19, pp. 5803–5808, May 2016.
- [15] J. Shi, H. Zhang, Z. Chen, L. Xu, and Z. Zhang, “A multi-functional nanoplatform for efficacy tumor theranostic applications,” *Asian J. Pharm. Sci.*, vol. 12, no. 3, pp. 235–249, May 2017.
- [16] T. Liu *et al.*, “Drug delivery with PEGylated MoS₂ nano-sheets for combined photothermal and chemotherapy of cancer,” *Adv. Mater.*, vol. 26, no. 21, pp. 3433–3440, Jun. 2014.
- [17] N. Li, W. Zhang, M. Khan, L. Lin, and J. M. Lin, “MoS₂-LA-PEI nanocomposite carrier for real-time imaging of ATP metabolism in glioma stem cells co-cultured with endothelial cells on a microfluidic system,” *Biosens. Bioelectron.*, vol. 99, pp. 142–149, Jan. 2018.
- [18] R. C. Rowe, P. J. Sheskey, and M. E. Quinn, *Handbook of pharmaceutical excipients*. Pharmaceutical Press, 2009.
- [19] A. Abuchowski, J. R. McCoy, N. C. Palczuk, T. van Es, and F. F. Davis, “Effect of covalent attachment of polyethylene glycol on immunogenicity and circulating life of bovine liver catalase.,” *J. Biol. Chem.*, vol. 252, no. 11, pp. 3582–3586, 1977.
- [20] A. Abuchowski, T. Van Es, N. C. Palczuk, and F. F. Davis, “Alteration of immunological properties of bovine serum albumin by covalent attachment of polyethylene glycol.,” *J. Biol. Chem.*, vol. 252, no. 11, pp. 3578–3581, 1977.
- [21] A. Ulman, *Langmuir–Blodgett Films*. 1991.
- [22] J. Kim, L. J. Cote, F. Kim, W. Yuan, K. R. Shull, and J. Huang, “Graphene oxide sheets at interfaces,” *J. Am. Chem. Soc.*, vol. 132, no. 23, pp. 8180–8186, Jun. 2010.
- [23] K. Vegso *et al.*, “Nonequilibrium phases of nanoparticle langmuir films,” *Langmuir*, vol. 28, no. 28, pp. 10409–10414, Jul. 2012.
- [24] I. G. Barnes G, *An Introduction to Interfacial Science*. Oxford University Press, 2005.
- [25] R. Reiter, H. Motschmann, H. Orendi, A. Nemetz, and W. Knoll, “Ellipsometric Microscopy. Imaging Monomolecular Surfactant Layers at the Air–Water Interface,” *Langmuir*, vol. 8, no. 7, pp. 1784–1788, Jul. 1992.
- [26] B. A. Noskov *et al.*, “Dynamic surface properties of C60-arginine and C60-L-lysine aqueous solutions,” *Colloids Surfaces A Physicochem. Eng. Asp.*, vol. 529, pp. 1–6, Sep. 2017.
- [27] X. W. Teng, X. Y. Liang, S. Rahman, and H. Yang, “Porous Nanoparticle Membranes: Synthesis and Application as Fuel-Cell Catalysts,” *Adv. Mater.*, vol. 17, no. 18, pp. 2237–2241, Sep. 2005.
- [28] M. Tahghighi, I. Mannelli, D. Janner, and J. Ignés-Mullol, “Tailoring plasmonic response by Langmuir–Blodgett gold nanoparticle templating for the fabrication of SERS substrates,” *Appl. Surf. Sci.*, vol. 447, pp. 416–422, 2018.
- [29] M. E. Villanueva, A. E. Lanterna, and R. V. Vico, “Hydrophobic silver nanoparticles interacting with phospholipids and stratum corneum mimic membranes in Langmuir monolayers,” *J. Colloid Interface Sci.*, vol. 543, pp. 247–255, 2019.
- [30] S. Datta, J. Biswas, and S. Bhattacharya, “How does spacer length of imidazolium gemini surfactants control the fabrication of 2D-Langmuir films of silver-nanoparticles at the air–water interface?,” *J. Colloid Interface Sci.*, vol. 430, pp. 85–92, Sep. 2014.
- [31] Y. Tian and J. H. Fendler, “Langmuir-Blodgett film formation from fluorescence-activated, surfactant-capped, size-selected CdS nanoparticles spread on water

- surfaces,” *Chem. Mater.*, vol. 8, no. 4, pp. 969–974, Jan. 1996.
- [32] M. Bardosova, M. E. Pemble, I. M. Povey, and R. H. Tredgold, “The langmuir-blodgett approach to making colloidal photonic crystals from silica spheres,” *Adv. Mater.*, vol. 22, no. 29, pp. 3104–3124, Aug. 2010.
- [33] M. Bodik *et al.*, “On the formation of hydrophobic carbon quantum dots Langmuir films and their transfer onto solid substrates,” *Diam. Relat. Mater.*, vol. 83, pp. 170–176, 2018.
- [34] K. Vegso *et al.*, “Silver nanoparticle monolayer-to-bilayer transition at the air/water interface as studied by the GISAXS technique: Application of a new paracrystal model,” *Langmuir*, vol. 28, no. 25, pp. 9395–9404, 2012.